



UNIVERSITI PUTRA MALAYSIA

**CHEMICAL CONSTITUENTS, BIOACTIVITY AND HPLC PROFILING OF
MICROWAVE-ASSISTED AND NORMAL EXTRACTION OF *Murraya*
*koenigii***

NOOR HASLIZAWATI BINTI ABU BAKAR

FS 2008 56

**CHEMICAL CONSTITUENTS, BIOACTIVITY AND HPLC PROFILING OF
MICROWAVE-ASSISTED AND NORMAL EXTRACTION OF *Murraya koenigii***

NOOR HASLIZAWATI BINTI ABU BAKAR

**MASTER OF SCIENCE
UNIVERSITI PUTRA MALAYSIA**

2008



Abstract of thesis presented to Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

**CHEMICAL CONSTITUENTS, BIOACTIVITY AND HPLC PROFILING OF
MICROWAVE-ASSISTED AND NORMAL EXTRACTION OF *Murraya koenigii***

By

NOOR HASLIZAWATI BINTI ABU BAKAR

June 2008

Chairman: Profesor Mohd. Aspollah Hj. Sukari, PhD

Faculty: Faculty of Science

Murraya koenigii (L.) Spreng is an aromatic plant, which is normally used as natural flavoring in curries and sources, and commonly found in India and Peninsular Asia. Locally, *Murraya koenigii* is known as curry leaf tree and one of the richest source of carbazole alkaloids. It has been reported that carbazole alkaloids possess various biological activities such as anti-tumor, anti-oxidative, anti-mutagenic and anti-inflammatory activities. In this work, six carbazole alkaloids: mahanimbine (**33**), girinimbine (**17**), murrayanine (**31**), murrayafoline-A (**62**), murrayacine (**39**) and 3-methylcarbazole (**63**), one long chain ester (**61**) together with β -sitosterol (**24**) were isolated from leaves, stem barks, and roots of *Murraya koenigii* by using normal extraction (NE) and microwave-assisted extraction (MAE). The structures of these compounds were established by spectroscopic method and by comparison with the previous reported works.

The essential oil of the fresh curry leaves was obtained using conventional hydrodistillation (CHD) and microwave-assisted hydrodistillation (MAHD), and were analyzed by gas chromatography and GC-MS techniques. Most of the components of the oils obtained from both techniques of distillation were rather similar but with different variations of quantities. In the essential oil obtained from conventional hydrodistillation (CHD), the major compound was α -farnesene (18.74%), whereas the major constituent of oils from microwave-assisted hydrodistillation (MAHD) was 4,11,11-trimethyl-8-methylenebicycloundec-4-ene (29.67%).

All crude extracts and carbazole alkaloids isolated from *Murraya koenigii* were screened their cytotoxic activities towards human T-promyelocytic leukemic cell lines (HL-60), human colon cancer cells (HT-29), human breast cancer cells (MCF-7) and human cervical carcinoma cancer cells (HeLa) and all isolated compounds were strongly active with IC_{50} values gave less than 20 μ g/ml. In the larvicidal activity, the crude extracts and isolated compounds were tested against the larvae of *Aedes aegypti*. All crude extracts and isolated compounds showed very strong activity against *Aedes aegypti* with LC_{50} values of between 0.68 ppm to 55.03 ppm. In the antimicrobial screening, only crude chloroform extract of roots of *Murraya koenigii* showed low activity against *Bacillus subtilis*, while the antifungal test showed that the crude chloroform extract of roots and murrayafoline-A (**62**) showed low activity against the *Candida albicans*.

The bioactivity tests carried out in this research which include antimicrobial activity of some pathogenic microbes, cytotoxicity tests against some cancer cell lines (HL-60, MCF-7, HT-29 and HeLa) and larvicidal activity properties against *Aedes aegypti* were

never been reported previously. In addition, the microwave-assisted extraction of the plant and the development of the profiling of the extracts based on using HPLC-UV technique were never been investigated before.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Master Sains

PEMENCILAN SEBATIAN KIMIA, AKTIVITI-AKTIVITI BIOLOGI DAN PEMPROFILAN KROMATOGRAFI CECAIR PRESTASI TINGGI BAGI PENGEKSTRAKAN GELOMBANG MIKRO DAN PENGEKSTRAKAN NORMAL DARIPADA *Murraya koenigii*

Oleh

NOOR HASLIZAWATI BINTI ABU BAKAR

Jun 2008

Pengerusi: Prof. Dr. Mohd. Aspollah Hj. Sukari, PhD

Fakulti: Fakulti Sains

Murraya koenigii (L.) Spreng adalah tumbuhan aromatik, yang biasanya di gunakan sebagai perisa semulajadi dalam masakan kari dan sebagainya, selalunya di jumpai di India dan Peninsular Asia. *Murraya koenigii* juga dikenali sebagai pokok daun kari dan merupakan salah satu sumber yang kaya dengan karbazol alkaloid. Karbazol alkaloid pernah dinyatakan bagi pelbagai aktiviti biologi antaranya ialah anti-kanser, anti-penuaan, anti-mutagenik dan anti-radang. Dalam kajian ini, enam karbazol alkaloid: mahanimbin (**33**), girinimbin (**17**), murrayanin (**31**), murrayafolin-A (**62**), murrayacin (**39**) dan 3-metilkarbazol (**63**), ester rantai panjang (**61**) bersama β -sitosterol (**24**) telah dipencilkan daripada bahagian daun, kulit batang dan akar *Murraya koenigii* dengan menggunakan kaedah pengekstrakan biasa (rendaman) (NE) dan kaedah pengekstrakan

menggunakan gelombang mikro (MAE). Kesemua sebatian telah dicirikan berdasarkan data spektroskopi dan perbandingan dengan data kajian sebelum ini.

Minyak pati daripada daun kari yang segar didapati dengan menggunakan kaedah penyulingan hidro (CHD) dan penyulingan hidro dengan gelombang mikro (MAHD), minyak pati di analisis menggunakan kaedah spektroskopi GC-MS. Kebanyakan komponen dalam minyak pati yang terhasil daripada kedua-dua teknik penyulingan tersebut adalah agak sama tetapi berbeza dari segi kuantiti. Sebatian utama minyak pati yang terhasil daripada penyulingan hidro (CHD) adalah α -farnesena (18.74%), dan bagi penyulingan hidro dengan gelombang mikro (MAHD) pula adalah 4,11,11-trimetil-8-metilenabisikloundek-4-ena (29.67%).

Kesemua ekstrak dan sebatian tulen daripada *Murraya koenigii* juga telah disaring dengan aktiviti sitotoksik terhadap sel kanser leukemia manusia (HL-60), sel kanser kolon manusia (HT-29), sel kanser payudara manusia (MCF-7) dan sel kanser servik manusia (HeLa), dan menunjukkan aktiviti yang tinggi dengan nilai IC_{50} adalah kurang daripada 20 μ g/mL. Dalam ujian aktiviti larva, kesemua ekstrak dan sebatian tulen telah diuji terhadap larva nyamuk Aedes (*Aedes aegypti*). Kesemua ekstrak dan sebatian tulen mempamerkan aktiviti yang tinggi terhadap larva *Aedes aegypti* dengan nilai LC_{50} adalah di antara 0.68 ppm hingga 55.03 ppm. Bagi penyaringan antimikrobial pula, hanya ekstrak klorofom daripada akar *Murraya koenigii* menunjukkan aktiviti yang rendah terhadap *Bacillus subtilis*, sementara dalam ujian antifungal ekstrak klorofom daripada akar dan murrayafoline-A aktiviti juga menunjukkan aktiviti yang rendah terhadap *Candida albicans*.

Penyelidikan ini diteruskan dengan mengkaji aktiviti biologi bagi aktiviti anti-mirob terhadap beberapa mikrob, ujian anti-sitotoksik melawan sel kanser (HL-60, MCF-7, HT-29 and HeLa) dan aktiviti larvicidal melawan *Aedes aegypti* kerana telah dikaji tiada laporan terdahulu mengenainya. Sebagai penambahan, teknik pengekstrakan gelombang mikro telah dibangunkan untuk pemprofilan bagi ekstrak menggunakan teknik HPLC-UV juga telah dikaji tiada penyelidikan mengenainya.

ACKNOWLEDGEMENT

In the name of Allah, most Gracious and most Merciful in giving me the strength and patience to complete this thesis.

I would like to express my highest gratitude and deepest appreciation to my supervisor, Professor Dr. Mohd Aspollah Hj. Sukari for his intellectual advices and suggestion throughout the development of this research. My sincere thanks and deepest gratitude is also extended to my supervisory committee members, Prof. Mawardi Rahmani, Prof. Kaida Khalid and En. Atan Md Sharif for their guidance and invaluable advices.

I wish to express my sincere gratitude to all the staff of Chemistry Department, especially En. Zainudin Samadi, En. Zainal Kassim, Puan Rusnani Amirudin, En. Abas Abd. Rahman and En. Johadi Iskandar for all their help and co-operation during this research. Immeasurable gratitude is also extended to the staff of Bioscience Institute (IBS) for their help and co-operation in bioassay screening.

My special thanks also goes to my laboratory mates, especially Nurul Waznah Muhd Sharif, Tang Sook Wah, Amy Yap Li Ching for their useful suggestions and encouragement throughout my research.

Grateful appreciation is also extended to my beloved family and friends, who have been very supportive throughout all these years.

I certify that an Examination Committee has met on 21st October 2008 to conduct the final examination of Noor Haslizawati binti Abu Bakar on her degree thesis entitled “Chemical constituents, Bioactivity and HPLC Profiling of Microwave-Assisted and Normal Extraction of *Murraya koenigii*” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the Degree of Master of Sciences.

Members of the Examination Committee were as follows:

Taufiq-Yap Yun Hin, PhD

Professor
Department of Chemistry
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Irmawati Ramli, PhD

Associate Professor
Department of Chemistry
Faculty of Science
Universiti Putra Malaysia
(Member)

Siti Mariam Md. Noor, PhD

Lecturer
Department of Chemistry
Faculty of Science
Universiti Putra Malaysia
(Member)

Faradiah, PhD

Associate Professor
Department of Chemistry
Faculty of Science
Universiti Kebangsaan Malaysia
(Member)

HASANAH MOHD. GHAZALI, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd. Aspollah Hj. Sukari, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Mawardi Rahmani, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Kaida Khalid, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Atan Md. Sharif, MSc

Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)

HASANAH MOHD. GHAZALI, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 12 February 2009

DECLARATION

I hereby declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

NOOR HASLIZAWATI ABU BAKAR

Date: 17 December 2009

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	v
ACKNOWLEDGEMENTS	viii
APPROVAL	ix
DECLARATION	xi
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xxiii
 CHAPTER	
 1 INTRODUCTION	
1.1 General	1
1.2 Natural Products Research in Malaysia	3
1.3 Rutaceae	4
1.3.1 Genus <i>Murraya</i>	6
1.3.2 <i>Murraya koenigii</i> (L.) Spreng	6
1.3.3 Uses and Medicinal properties	8
1.3.4 Essential oils	9
1.4 Bioassays	12
1.4.1 Cancer and Cytotoxic drugs	12
1.4.2 Larvicidal activity	13
1.5 Microwave-assisted Extraction (MAE)	14
1.6 High Performance Liquid Chromatography (HPLC)	17
1.7 Problem Statement	18
1.8 Objectives	19
 2 LITERATURE REVIEW	
2.1 Phytochemical studies on <i>Murraya</i>	20
2.2 Previous works on <i>Murraya koenigii</i>	26
2.2.1 Phytochemical studies on <i>Murraya koenigii</i>	26
2.2.2 Biological activity on <i>Murraya koenigii</i>	33
2.2.3 Essential oils of <i>Murraya koenigii</i>	38
2.3 Previous works on Microwave-assisted extraction	40
2.4 Previous works on HPLC Profiling	41
 3 MATERIALS AND METHODS	
3.1 Instrument	43
3.1.1 Melting point apparatus	43
3.1.2 Infrared Spectroscopy (IR)	43
3.1.3 Ultraviolet Visible Spectroscopy (UV-Vis)	43
3.1.4 Gas Chromatography-Mass Spectrometry (GCMS)	43
3.1.5 Nuclear Magnetic Resonance (NMR)	44
3.1.6 High Performance Liquid Chromatography (HPLC)	44
3.1.7 Microwave Extraction System (MES)	45



3.2	Chromatographic Method	45
3.2.1	Column Chromatography (CC)	45
3.2.2	Thin Layer Chromatography (TLC)	45
3.3	Experimental Method	46
3.3.1	Plant material	46
3.3.2	Extraction	46
3.3.3	Separation and Purification	47
3.4	Isolation of Chemical constituents from Leaves of <i>Murraya koenigii</i>	50
3.4.1	Nor mal extraction (NE)	50
3.4.2	Microwave-assisted extraction (MAE)	51
3.5	Isolation of Chemical constituents from Stem barks of <i>Murraya koenigii</i>	53
3.5.1	Nor mal extraction (NE)	53
3.5.2	Microwave-assisted extraction (MAE)	56
3.6	Isolation of chemical constituents from Roots of <i>Murraya koenigii</i>	57
3.6.1	Nor mal extraction (NE)	57
3.6.2	Microwave-assisted extraction (MAE)	61
3.7	Essential oils of Fresh Curry leaves	62
3.7.1	Plant material	62
3.7.2	Conventional Hydrodistillation (CHD)	63
3.7.3	Microwave-assisted Hydrodistillation (MAHD)	63
3.7.4	Isolation of the oils	64
3.8	Profiling Method	67
3.8.1	Sample preparation	67
3.8.3	HPLC profiling	67
3.9	Bioassay Screening Method	68
3.9.1	Cytotoxic screening	68
3.9.2	Antimicrobial activity	69
3.9.3	Larvicidal activity	71

4 RESULTS AND DISCUSSION

4.1	Bioactive constituents from <i>Murraya koenigii</i> with Normal extraction (NE) and Microwave-assisted extraction (MAE) techniques	72
4.2	Extraction and Isolation of chemical constituents from Leaves of <i>Murraya koenigii</i>	77
4.2.1	Characterization of Mahanimbine (33)	78
4.2.2	Characterization of Ethyl octadecanoate (61)	97
4.3	Extraction and Isolation of chemical constituents from Stem barks of <i>Murraya koenigii</i>	104
4.3.1	Characterization of Grinimbine (17)	105
4.3.2	Characterization of Murrayacine (39)	124
4.3.3	Characterization of Murrayanine (31)	140
4.4	Extraction and Isolation of chemical constituents from Roots	

of <i>Murraya koenigii</i>	157
4.4.1 Characterization of Murrayafoline-A (62)	157
4.4.2 Characterization of 3-Methylcarbazole (63)	173
4.4.3 Characterization of β -Sitosterol (24)	187
4.5 Characterization of chemical compositions of essential oil obtained by Conventional Hydrodistillation (CHD) and Microwave-assisted Hydrodistillation (MAHD)	193
4.6 High Performance Liquid Chromatography (HPLC) profiling on crude extracts of <i>Murraya koenigii</i> obtained by normal extraction (NE) and Microwave-assisted extraction (MAE)	205
4.6.1 HPLC profiling on crude leaves extracts	206
4.6.2 HPLC profiling on crude stem barks extracts	210
4.6.3 HPLC profiling on crude roots extracts	214
4.7 Bioassay Screening	218
4.7.1 Cytotoxic screening	218
4.7.2 Larvicidal activity	221
4.7.3 Antimicrobial and Antifungal test	224
5 CONCLUSIONS	227
REFERENCES	230
APPENDICES	237
BIODATA OF STUDENT	281
LIST OF PUBLICATIONS	282

LIST OF TABLES

Table	Page
4.1 The weight and percentage yield from leaves extract of <i>Murraya koenigii</i>	72
4.2 The weight and percentage yield from stem barks extract of <i>Murraya Koenigii</i>	72
4.3 The weight and percentage yield from roots extract of <i>Murraya koenigii</i>	73
4.4 ¹ H NMR (400 MHz) spectral data for mahanimbine (33)	81
4.5 ¹³ C NMR (100 MHz) spectral data for mahanimbine (33)	82
4.6 2D-NMR spectral data for mahanimbine (33)	83
4.7 ¹ H NMR (400 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data for compound ethyl octadecanoate (61)	99
4.8 ¹ H NMR (400 MHz) spectral data for girinimbine (17)	107
4.9 ¹³ C NMR (100 MHz) spectral data for girinimbine (17)	108
4.10 2D-NMR spectral data for girinimbine (17)	109
4.11 ¹ H NMR (400 MHz) spectral data for murrayacine (39)	126
4.12 ¹³ C NMR (100 MHz) spectral data for murrayacine (39)	126
4.13 2D-NMR spectral data for murrayacine (39)	127
4.14 ¹ H NMR (400 MHz) spectral data for murrayanine (31)	142
4.15 ¹³ C NMR (100 MHz) spectral data for murrayanine (31)	142
4.16 2D-NMR spectral data for murrayanine (31)	143
4.17 ¹ H NMR (400 MHz) spectral data for murrayafoline-A (62)	159
4.18 ¹³ C NMR (100 MHz) spectral data for murrayafoline-A (62)	160
4.19 2D-NMR spectral data for murrayafoline-A (62)	160
4.20 ¹ H NMR (400 MHz) spectral data for 3-methylcarbazole (63)	174



4.21	¹³ C NMR (100 MHz) spectral data for 3-methylcarbazole (63)	175
4.22	2D-NMR spectral data for 3-methylcarbazole (63)	175
4.23	Weight, percentage yield and physical appearance of essential oil obtained from fresh Curry leaves.	193
4.24	The major constituents of essential oils from conventional hydrodistillation (CHD).	194
4.25	The major constituents of essential oils from microwave-assisted hydrodistillation (MAHD).	195
4.26	Composition of essential oil obtained from microwave-assisted hydrodistillation (MAHD) of curry leaves.	198
4.27	Composition of essential oil obtained from conventional hydrodistillation of curry leaves.	201
4.28	Cytotoxic activity of crude extracts of <i>Murraya koenigii</i> against HL-60, MCF-7, HT-29 and HeLa cancer cell lines.	218
4.29	Cytotoxic activity of pure compounds of <i>Murraya koenigii</i> against HL-60, MCF-7, HT-29 and HeLa cancer cell lines.	219
4.30	The larvicidal activity of crude extracts from <i>Murraya koenigii</i> against mosquito larvae of <i>Aedes aegypti</i> .	223
4.31	The larvicidal activity of pure compounds from <i>Murraya koenigii</i> against mosquito larvae of <i>Aedes aegypti</i> .	223
4.32	Antimicrobial activity test of crude extracts and essential oils of <i>Murraya koenigii</i> .	225
4.33	Antimicrobial activity test of pure compounds of <i>Murraya koenigii</i> .	225
4.34	Antifungal activity test of crude extracts and essential oils of <i>Murraya koenigii</i> .	226
4.35	Antifungal activity test of pure compounds of <i>Murraya koenigii</i> .	226

LIST OF FIGURES

Figure	Page
1.1 The leaves of <i>Murraya koenigii</i> (L.) spreng.	11
1.2 The stem barks of <i>Murraya koenigii</i> (L.) spreng.	11
1.3 The roots of <i>Murraya koenigii</i> (L.) spreng	11
3.1 Flow chart for the extraction and isolation of <i>Murraya koenigii</i>	48
3.2 Microwave extraction-Milestone ETHOS SEL MicrowaveLabstation bath reactor	49
3.3 Flow chart for the extraction of essential oil of <i>Murraya koenigii</i>	65
3.4 Pyrex or corning glass Dean and Stark apparatus	66
3.5 Microwave-assisted Hydrodistillation (MAHD)	66
4.1 Scheme for the isolation of the leaves of <i>Murraya koenigii</i> using normal extraction.	74
4.2 Scheme for the isolation of the stem barks of <i>Murraya koenigii</i> using normal extraction.	74
4.3 Scheme for the isolation of the roots of <i>Murraya koenigii</i> using normal extraction.	75
4.4 Scheme for the isolation of the leaves of <i>Murraya koenigii</i> using microwave-assisted extraction.	76
4.5 Scheme for the isolation of the stem barks of <i>Murraya koenigii</i> using microwave assisted extraction.	76
4.6 Scheme for the isolation of the roots of <i>Murraya koenigii</i> using microwave-assisted extraction.	84
4.7 Mass fragmentation pattern of mahanimbine (33)	84
4.8 IR spectrum of mahanimbine (33)	85
4.9 Mass spectrum of mahanimbine (33)	86
4.10a ¹ H NMR spectrum of mahanimbine (33)	87

4.10b	Expanded ^1H NMR spectrum of mahanimbine (33)	88
4.11a	^{13}C NMR spectrum of mahanimbine (33)	89
4.11b	Expanded ^{13}C NMR spectrum of mahanimbine (33)	90
4.12	DEPT spectrum of mahanimbine (33)	91
4.13a	COSY spectrum of mahanimbine (33)	92
4.13b	Expanded COSY spectrum of mahanimbine (33)	93
4.14a	HMQC spectrum of mahanimbine (33)	94
4.14b	Expanded HMQC spectrum of mahanimbine (33)	95
4.15a	HMBC spectrum of mahanimbine (33)	96
4.15b	Expanded HMBC spectrum of mahanimbine (33)	97
4.16	IR spectrum of ethyl octadecanoate (61)	100
4.17	MS spectrum of ethyl octadecanoate (61)	101
4.18	^1H NMR spectrum of ethyl octadecanoate (61)	102
4.19	^{13}C NMR spectrum of ethyl octadecanoate (61)	103
4.20	Mass fragmentation pattern of girinimbine (17)	110
4.21	IR spectrum of girinimbine (17)	111
4.22	Mass spectrum of girinimbine (17)	112
4.23a	^1H NMR spectrum of girinimbine (17)	113
4.23b	Expanded ^1H NMR spectrum of girinimbine (17)	114
4.24a	^{13}C NMR spectrum of girinimbine (17)	115
4.24b	Expanded ^{13}C NMR spectrum of girinimbine (17)	116
4.25	DEPT spectrum of girinimbine (17)	117
4.26a	COSY spectrum of girinimbine (17)	118
4.26b	Expanded COSY spectrum of girinimbine (17)	119

4.27a	HMQC spectrum of girinimbine (17)	120
4.27b	Expanded HMQC spectrum of girinimbine (17)	121
4.28a	HMBC spectrum of girinimbine (17)	122
4.28b	Expanded HMBC spectrum of girinimbine (17)	123
4.29	IR spectrum of murrayacine (39)	128
4.30	MS spectrum of murrayacine (39)	129
4.31a	^1H NMR spectrum of murrayacine (39)	130
4.31b	Expanded ^1H NMR spectrum of murrayacine (39)	131
4.32	^{13}C NMR spectrum of murrayacine (39)	132
4.33	DEPT spectrum of murrayacine (39)	133
4.34a	COSY spectrum of murrayacine (39)	134
4.34b	Expanded COSY spectrum of murrayacine (39)	135
4.35a	HMQC spectrum of murrayacine (39)	136
4.35b	Expanded HMQC spectrum of murrayacine (39)	137
4.36a	HMBC spectrum of murrayacine (39)	138
4.36b	Expanded HMBC spectrum of murrayacine (39)	139
4.36	Mass fragmentation pattern of murrayanine (31)	144
4.38	IR spectrum of murrayanine (31)	145
4.39	MS spectrum of murrayanine (31)	146
4.40a	^1H NMR spectrum of murrayanine (31)	147
4.40b	Expanded ^1H NMR spectrum of murrayanine (31)	148
4.41	^{13}C NMR spectrum of murrayanine (31)	149
4.42	DEPT spectrum of murrayanine (31)	150
4.43a	COSY spectrum of murrayanine (31)	151

4.43b	Expanded COSY spectrum of murrayanine (31)	152
4.44a	HMQC spectrum of murrayanine (31)	153
4.44b	Expanded HMQC spectrum of murrayanine (31)	154
4.45a	HMBC spectrum of murrayanine (31)	155
4.45b	Expanded HMBC spectrum of murrayanine (31)	156
4.46	IR spectrum of murrayafoline-A (62)	161
4.47	MS spectrum of murrayafoline-A (62)	162
4.48a	^1H NMR spectrum of murrayafoline-A (62)	163
4.48b	Expanded ^1H NMR spectrum of murrayafoline-A (62)	164
4.49	^{13}C NMR spectrum of murrayafoline-A (62)	165
4.50	DEPT spectrum of murrayafoline-A (62)	166
4.51a	COSY spectrum of murrayafoline-A (62)	167
4.51b	Expanded COSY spectrum of murrayafoline-A (62)	168
4.52a	HMQC spectrum of murrayafoline-A (62)	169
4.52b	Expanded HMQC spectrum of murrayafoline-A (62)	170
4.53a	HMBC spectrum of murrayafoline-A (62)	171
4.53b	Expanded HMBC spectrum of murrayafoline-A (62)	172
4.54	IR spectrum of 3-methylcarbazole (63)	176
4.55	MS spectrum of 3-methylcarbazole (63)	177
4.56a	^1H NMR spectrum of 3-methylcarbazole (63)	178
4.56b	Expanded ^1H NMR spectrum of 3-methylcarbazole (63)	179
4.57	^{13}C NMR spectrum of 3-methylcarbazole (63)	180
4.58	DEPT spectrum of 3-methylcarbazole (63)	181
4.59a	COSY spectrum of 3-methylcarbazole (63)	182

4.59b	Expanded COSY spectrum of 3-methylcarbazole (63)	183
4.60	HMQC spectrum of 3-methylcarbazole (63)	184
4.61a	HMBC spectrum of 3-methylcarbazole (63)	185
4.61b	Expanded HMBC spectrum of 3-methylcarbazole (63)	186
4.62	IR spectrum of β -sitosterol (24)	189
4.63	MS spectrum of β -sitosterol (24)	190
4.64	^1H NMR spectrum of β -sitosterol (24)	191
4.65	^{13}C NMR spectrum of β -sitosterol (24)	192
4.66	GC chromatogram for curry leaves essential oils obtained from microwave-assisted hydrodistillation (MAHD)	197
4.67	GC chromatogram for curry leaves essential oils obtained from conventional hydrodistillation (CHD)	200
4.68	HPLC chromatograms of hexane extracts from <i>Murraya koenigii</i> leaves obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), and murrayanine (3), each concentration was 1 mg/mL.	207
4.69	HPLC chromatograms of chloroform extracts from <i>Murraya koenigii</i> leaves obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1) and girinimbine (2), each concentration was 1 mg/mL.	208
4.70	HPLC chromatograms of methanol extracts from <i>Murraya koenigii</i> leaves obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1) and girinimbine (2), each concentration was 1 mg/mL.	209
4.71	HPLC chromatograms of hexane extracts from <i>Murraya koenigii</i> stem barks obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), and murrayanine (3), each concentration was 1 mg/mL.	211

- 4.72 HPLC chromatograms of chloroform extracts from *Murraya koenigii* stem barks obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), and murrayanine (3), each concentration was 1 mg/mL. 212
- 4.73 HPLC chromatograms of methanol extracts from *Murraya koenigii* stem barks obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), and murrayanine (3), each concentration was 1 mg/mL. 213
- 4.74 HPLC chromatograms of hexane extracts from *Murraya koenigii* roots obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), murrayanine (3) and murayafoline-A (4), each concentration was 1 mg/mL. 215
- 4.75 HPLC chromatograms of chloroform extracts from *Murraya koenigii* roots obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), murrayanine (3) and murayafoline-A (4), each concentration was 1 mg/mL. 216
- 4.76 HPLC chromatograms of methanol extracts from *Murraya koenigii* roots obtained from normal extraction (NE) and microwave-assisted extraction (MAE). Isolated compounds used were mahanimbine (1), girinimbine (2), murrayanine (3) and murayafoline-A (4), each concentration was 1 mg/mL. 217

LIST OF ABBREVIATIONS

α	Alpha
β	Beta
δ	Chemical shift in ppm
^{13}C	Carbon-13
CHCl_3	Chloroform
$^{\circ}\text{C}$	Degree in Celcius
CDCl_3	Deutrated chloroform
COSY	Correlated spectroscopy
cm	Centimeter
J	Coupling constant in Hertz
d	Doublet
DEPT	Distortionless Enhancement by Polarisation Transfer
DMSO	Dimethylsulfoxide
EIMS	Electron Impact-MASS spectroscopy
EA	Ethyl Acetate
G	Gram
GC	Gas Chromatography
GC-MS	Gas Chromatography-Mass Spectroscopy
^1H	Proton
HMBC	Heteronuclear Mutiple Bond Connectivity
HMQC	Heteronuclear Multiple Quantum Correlation
HPLC	High Performance Liquid Chromatography
Hz	Hertz
OH	Hydroxy
IC	Inhibition Concentration
IR	Infrared
LC	Lethal concentration
m/z	Mass per charge
MS	Mass spectroscopy
MeOH	Methanol



OMe	Methoxy
Me	Methyl
m.p.	Melting point
mL	Milliliter
mm	Milimeter
μg	Microgram
μL	Microliter
MAE	Microwave-assisted extraction
mg	Milligram
M ⁺	Molecular ion
<i>m</i>	Multiplet
nm	Nanometer
NMR	Nuclear Magnetic Resonance
ppm	Part per million
KBr	Potassium bromide
<i>s</i>	Singlet
<i>t</i>	Triplet
TLC	Thin layer chromatography
UV	Ultraviolet
WHO	World Health Organization